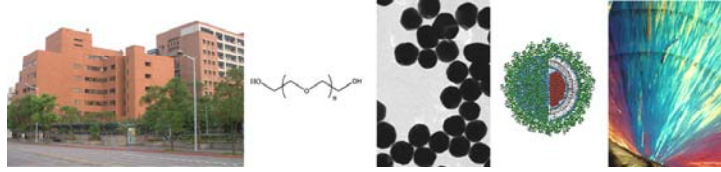


Anti-PEG



Sandwich ELISA assay (AGP4 coating / 3.3-biotin detection)

Materials:

- EIA plate (Nunc. Maxisorp 430341)
- Coating buffer (5.3 g Na_2CO_3 + 4.2 g NaHCO_3 /liter, pH=8.0, adjust pH with 12N HCl)
- PBS (0.14 M NaCl, 2.7 mM KCl, 1.5 mM KH_2PO_4 , 8.1 mM Na_2HPO_4 , pH 7.4)
- Blocking solution: 5% skim milk (Difco #232100) in PBS
- Dilution buffer: 2% skim milk in PBS.
- Washing buffer: PBS and PBS-T (PBS containing 0.05% Tween-20)
- Streptavidin-HRP (Jackson ImmunoResearch #016-030-084)
- HRP substrate: 50 mg/100 ml ABTS (Sigma #A-1888) in 100 mM phosphate-citrate buffer pH 4.0 (17.4 g K_2HPO_4 , 21 g citric acid in 1 Liter Q- H_2O). Immediately before use, add 3 μl of 30% H_2O_2 per 10 ml ABTS substrate solution.

Procedure:

- Dilute AGP4 to 5 $\mu\text{g}/\text{ml}$ in coating buffer. You need 5 ml per microtiter plate.
- Add 50 μl diluted AGP4 per well and incubate at 37°C for 4 h and then at 4°C overnight.
- Wash plates 3X with PBS.
- Add 200 μl blocking solution per well for 2 hours at room temperature.
- Dilute PEG-compound in dilution buffer to suitable concentrations
- Wash wells 3X with PBS
- Add graded concentrations of PEG-compound (50 $\mu\text{l}/\text{well}$) and incubate 2 h at room temperature.
- Wash with PBS-T 3X and PBS 2X.
- Add 50 $\mu\text{l}/\text{well}$ 3.3-biotin (5 $\mu\text{g}/\text{ml}$ in dilution buffer) for 1 h at room temperature.
- Wash wells with PBS-T 3X and with PBS 2X.
- Add 50 $\mu\text{l}/\text{well}$ streptavidin-HRP (1 $\mu\text{g}/\text{ml}$ in dilution buffer) for 1 h at room temperature
- Wash wells with PBS-T 6X and with PBS 2X.
- Add 100 $\mu\text{l}/\text{well}$ freshly prepared ABTS substrate for 30 min in dark at room temperature.
- Read absorbance of the wells at 405 nm.