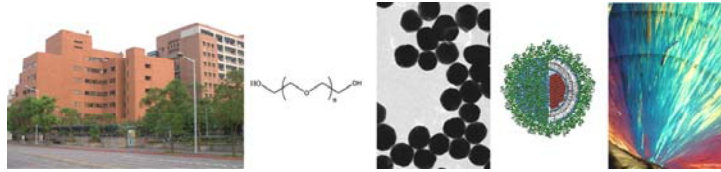


# Anti-PEG



## Sandwich ELISA assay (AGP4 coating / AGP3-biotin or AGP4-biotin detection)

### Materials:

- EIA plate (Nunc. Maxisorp 430341)
- Coating buffer (5.3 g  $\text{Na}_2\text{CO}_3$  + 4.2 g  $\text{NaHCO}_3$  /liter, pH=8.0, adjust pH with 12N HCl)
- PBS (0.14 M NaCl, 2.7 mM KCl, 1.5 mM  $\text{KH}_2\text{PO}_4$ , 8.1 mM  $\text{Na}_2\text{HPO}_4$ , pH 7.4)
- Blocking solution: 5% skim milk (Difco #232100) in PBS
- Dilution buffer: 2% skim milk in PBS.
- Washing buffer: PBS and PBS-T (PBS containing 0.05% Tween-20)
- Streptavidin-HRP (Jackson ImmunoResearch #016-030-084)
- HRP substrate: 50 mg/100 ml ABTS (Sigma #A-1888) in 100 mM phosphate-citrate buffer pH 4.0 (17.4 g  $\text{K}_2\text{HPO}_4$ , 21 g citric acid in 1 Liter Q- $\text{H}_2\text{O}$ ). Immediately before use, add 3  $\mu\text{l}$  of 30%  $\text{H}_2\text{O}_2$  per 10 ml ABTS substrate solution.

### Procedure:

- Dilute AGP4 to 5  $\mu\text{g}/\text{ml}$  in coating buffer. You need 5 ml per microtiter plate.
- Add 50  $\mu\text{l}$  diluted AGP4 per well and incubate at 37°C for 4 h and then at 4°C overnight.
- Wash plates 3X with PBS.
- Add 200  $\mu\text{l}$  blocking solution per well for 2 hours at room temperature.
- Dilute PEG-compound in dilution buffer to suitable concentrations
- Wash wells 3X with PBS
- Add graded concentrations of PEG-compound (50  $\mu\text{l}/\text{well}$ ) and incubate 2 h at room temperature.
- Wash with PBS-T 3X and PBS 2X.
- Mix 5  $\mu\text{g}/\text{ml}$  AGP4-biotin or AGP3-biotin with 1  $\mu\text{g}/\text{ml}$  streptavidin-HRP in dilution buffer.
- Add 50  $\mu\text{l}/\text{well}$  mixture for 1 h at room temperature.
- Wash wells with PBS-T 6X and with PBS 2X.
- Add 100  $\mu\text{l}/\text{well}$  freshly prepared ABTS substrate for 30 min in dark at room temperature.
- Read absorbance of the wells at 405 nm.